

BARBIGERONE, A NEW PYRANOISOFLAVONE FROM SEEDS OF *TEPHROSIA BARBIGERA*

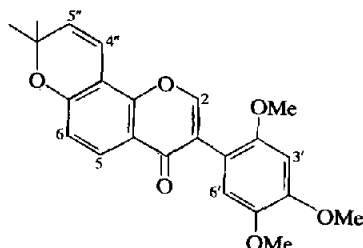
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Key Word Index—*Tephrosia barbiger*; Leguminosae; barbigerone; 2',4',5'-trimethoxy-6'',6''-dimethylpyrano(2'',3'':7,8)isoflavone.

The genus *Tephrosia* is known to be rich in flavonoids, isoflavonoids and rotenoids of complex structures [1–6]. In the present study, a new isoflavone, barbigerone (1), was found as the major flavonoid of the seeds of *Tephrosia barbiger*, an annual species widespread in Zaïre.



1 Barbigerone

The compound analysed for $C_{23}H_{22}O_6$, M^+ 394,1417 ($C_{23}H_{22}O_6$ requires M^+ 394,1416). UV λ_{max}^{MeOH} nm(log ϵ): 236(4.35), 263(4.47), 294(4.08) and 323(sh 3.86) which remained unaffected on the addition of NaOAc or $AlCl_3$. NMR(100 MHz, $CDCl_3$; δ , TMS internal standard): a large singlet at 1.48 (6H, Me_2C); three sharp singlets at 3.75, 3.84 and 3.91 (3H each, 2',4',5'-triMeO-); four one-proton doublets (which are typical of the indicated chromene ring system [7–11]) at 5.68 (H-5'', $J=10$ Hz), 6.82 (H-4'', $J=10$ Hz), 6.84 (H-6, $J=9$ Hz) and 8.06 (H-5, $J=9$ Hz); the two one-proton singlets at 6.62 (H-3') and 6.97 (H-6') indicate unequivocally a 2',4',5'-trioxygenation pattern for the side phenyl nucleus; finally, the sharp singlet at 7.96 (1H, H-2) which is a characteristic feature of isoflavones [13]. The MS showed prominent peaks at m/e (rel. int.) 394 (M^+ , 100), 379 (88), 363 (42), 349 (26), 203 (14), 191 (19), 189,5 (31), 187 (19), 182 (19), 168 (26), 149 (11), which was consistent with the proposed structure.

Barbigerone is therefore a novel substance whose structure is closely related to that of the rotenoid, deguelin, which belongs to the restricted group of angular 5-deoxypyranoisoflavones. Only four other compounds of this type are known in nature; all from the Leguminosae: jamaicin (2'-methoxy-4',5'-methylenedioxy-6'',6''-dimethylpyrano(2'',3'':7,8)isoflavone) from *Piscidia erythrina* [7, 13], ferrugone (2',5'-dimethoxy-3',4'-methylenedioxy-6'',6''-dimethylpyrano(2'',3'':7,8)isoflavone) from *Milletia ferruginea*

[8] and calopogoniumisoflavones A and B (respectively: 4'-methoxy- and 3',4'-methylenedioxy-6'',6''-dimethylpyrano(2'',3'':7,8)isoflavone) from *Calopogonium muconoides* [9–11].

EXPERIMENTAL

Seeds of *Tephrosia barbiger* L. were collected from Kinshasa's Plateaus, Zaïre. The whole, dried, finely pulverized seeds (110 g) were successively extracted in a Soxhlet with petrol (30–80°), Et_2O and EtOH. After evapn, the petrol extract (6.5 g) afforded a yellow solid (150 mg) which was purified by column chromatography over Si gel (Me_2CO gradually added to C_6H_6) and cryst. (25 mg) from abs. EtOH. The Et_2O and EtOH extract concentrates (1.5 and 3.9 g, respectively), when submitted to analytical TLC (toluene– Me_2CO , 4:1), showed almost identical compositions. Prep. TLC of these mixtures afforded several products but the main constituent (120 mg) was identical with that isolated from petrol: mp 153–4°, mmp undepressed, analytical TLC and co-TLC (R_f 0.51). This compound appeared as a dark spot in UV light (254 nm), gave an intense yellow colour with 5% H_2SO_4 –EtOH spray after 10 min at 100° but no ferric chloride reaction.

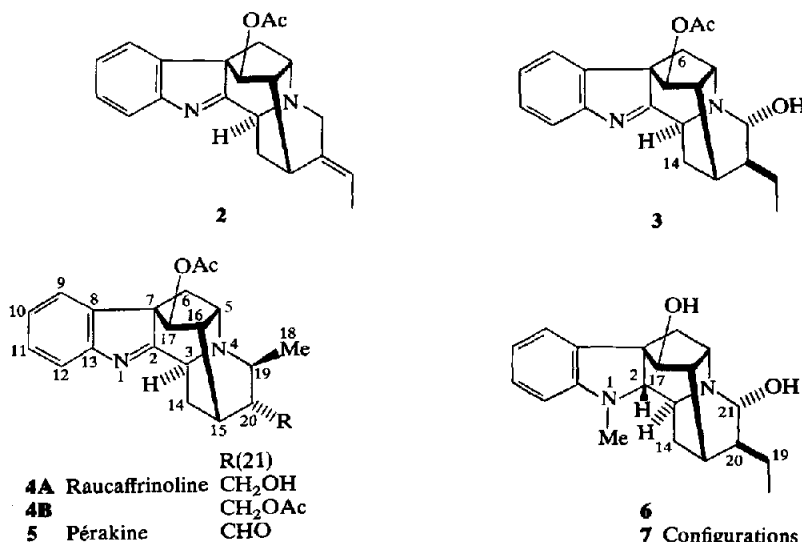
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REFERENCES

- Vleggar, R., Smalberger, T. M. and Dewaal, H. L. (1972) *Tetrahedron Letters* 703.
- Smalberger, T. M., Van Den Berg, A. J. and Vleggar, R. (1973) *Tetrahedron* **29**, 3099.
- Smalberger, T. M., Vleggar, R. and Weber, J. C. (1974) *Tetrahedron* **30**, 3927.
- Smalberger, T. M., Vleggar, R. and Weber, J. C. (1975) *Tetrahedron* **31**, 2297.
- Sarin, J. P. S., Singh, S., Garg, H. S., Khanna, N. M. and Dhar, M. M. (1976) *Phytochemistry* **15**, 232.
- Krupadanam, G. L. D., Sarma, P. N., Srimannarayana, G. and Subba Rao, N. V. (1977) *Tetrahedron Letters* 2125.

7. Schwarz, J. S. P., Cohen, A. I., Ollis, W. D., Kaczka, E. A. and Jackman, L. M. (1964) *Tetrahedron* **20**, 1317.
8. Highet, R. J. and Highet, P. F. (1967) *J. Org. Chem.* **32**, 1055.
9. Vilain, C. and Jadot, J. (1975) *Bull. Soc. R. Sci. Liège* **44**, 306.
10. Vilain, C. and Jadot, J. (1976) *Bull. Soc. R. Sci. Liège* **45**, 468.
11. Vilain C. (1978) Doctoral dissertation, Liège.
12. Markham, K. R. and Mabry, T. J. (1975) in *The Flavonoids* (Harborne J. B., Mabry T. J. and Mabry H., eds.). Chapman & Hall, London.
13. Stamm, O. A., Schmid, H. and Buchi, J. (1958) *Helv. Chim. Acta* **41**, 2006.

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7 Configurations inverses en 20, 21